

# **The application of human periodontal ligament stem cells and biomimetic silk scaffold for in situ tendon regeneration**

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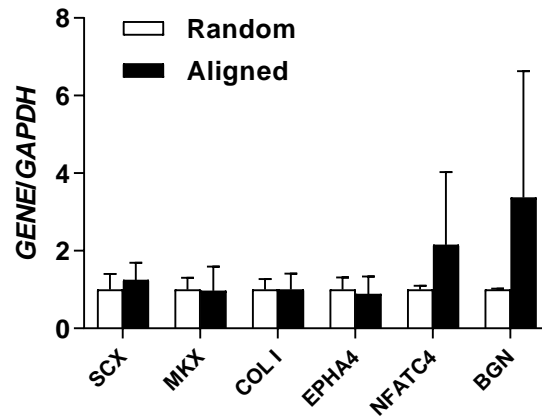
## Supplementary methods

### Tenogenic differentiation of hPDLSCs cultured on silk scaffolds

Human PDLSCs were seeded on random and aligned silk scaffolds, and cultured in high glucose DMEM (Gibco, C11995500) supplemented with 10% fetal bovine serum (Wisent, 086-550), 1% penicillin-streptomycin (Gibco, 15140122) and 50 µg/ml L-ascorbic acid 2-phosphate (A2-P; Sigma-Aldrich, A8960). After 7 days of culture, RNA extraction (Tiangen, DP430), cDNA reverse transcription (Toyobo, FSQ-201) and quantitative polymerase chain reaction (qPCR; Agbio, AG11718) were performed according to the manufacturers' protocols. All primers used in this study were summarized in Table S1. Representative results are displayed as target gene expression normalized to housekeeping gene GAPDH.

**Table S1.** Primers used for qPCR.

Genes	5'-3'	Primers
Glyceraldehyde 3-phosphate dehydrogenase ( <i>GAPDH</i> )	Forward	TGACGCTGGGGCTGGCATTG
	Reverse	GGCTGGTGGTCCAGGGGTCT
Scleraxis ( <i>SCX</i> )	Forward	CGAGAACACCCAGCCCAAAC
	Reverse	CTCCGAATCGCAGTCTTTCTGTC
Mohawk ( <i>MKX</i> )	Forward	GAAGGCAACTTTGTCTATCGCA
	Reverse	TGATCTCCTTCCAATACGTGTC
Collagen type I ( <i>COL I</i> )	Forward	CGATGGATTCCAGTTCGAGTAT
	Reverse	CATCGACAGTGACGCTGTAGG
EPH receptor A4 ( <i>EPHA4</i> )	Forward	AGTGGGCTGTGACAATCTGGAATA
	Reverse	CATTTAGACGGAAGTGAAGAGGGT
Nuclear factor of activated T-cells 4 ( <i>NFATC4</i> )	Forward	AAGGGTGAGACGGACATCG
	Reverse	CCGCCCATTGGAGACATAA
Biglycan ( <i>BGN</i> )	Forward	GATGGCCTGAAGCTCAA
	Reverse	GGTTTGTGAAGAGGCTG



**Figure S1.** Tenogenic differentiation of hPDLSCs cultured on silk scaffolds. Human PDLSCs were cultured on random and aligned silk scaffolds for 7 days. Gene expression of tendon-related genes was evaluated by qPCR. Expression levels of the random group were set as 1 in the quantified data. No significant difference was found ( $p \geq 0.05$ ).